

Life Sciences Reporting Summary

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For further information on the points included in this form, see [Reporting Life Sciences Research](#). For further information on Nature Research policies, including our [data availability policy](#), see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

► Experimental design

1. Sample size

Describe how sample size was determined.

G power software was used to calculate the sample size.

2. Data exclusions

Describe any data exclusions.

No data was excluded.

3. Replication

Describe whether the experimental findings were reliably reproduced.

experimental findings were reliably reproduced

4. Randomization

Describe how samples/organisms/participants were allocated into experimental groups.

The mice were allocated to groups randomly.

5. Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis.

No blinding was used.

Note: all studies involving animals and/or human research participants must disclose whether blinding and randomization were used.

6. Statistical parameters

For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or the Methods section if additional space is needed).

n/a Confirmed

- ☐ ☒ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
- ☐ ☒ A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly.
- ☐ ☒ A statement indicating how many times each experiment was replicated
- ☐ ☒ The statistical test(s) used and whether they are one- or two-sided (note: only common tests should be described solely by name; more complex techniques should be described in the Methods section)
- ☐ ☒ A description of any assumptions or corrections, such as an adjustment for multiple comparisons
- ☐ ☒ The test results (e.g. p values) given as exact values whenever possible and with confidence intervals noted
- ☐ ☒ A summary of the descriptive statistics, including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
- ☐ ☒ Clearly defined error bars

See the web collection on [statistics for biologists](#) for further resources and guidance.

► Software

Policy information about [availability of computer code](#)

7. Software

Describe the software used to analyze the data in this study.

Flowjo, Integrative Genomics Viewer, Prism

► Materials and reagents

Policy information about [availability of materials](#)

8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a for-profit company.

RORgtM/M mice are available from the authors.

9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

Antigen	Source	clone	Cat.	Lot	Species	Application
IL-17A	eBioscience	eBio17B7	17-7177-81	E026271	mouse, rat	FC
CD4	Biologend	GK1.5	100406	B225057	mouse	FC
CD8	eBioscience	53-6.7	15-0081-82	E026285	mouse	FC
Thy1.2	eBioscience	53-2.1	12-0902-81	E01379-1634	mouse	FC
CD24	eBioscience	M1/69	15-0242-81	E031467	mouse	FC
CD3	BD Biosciences	145-2C11	553066	06163	mouse	FC
CD19	eBioscience	eBio1D3	15-0193-81	E06107-1631	mouse	FC
CD11b	eBioscience	M1/70	12-0112-82	E01073-150	mouse	FC
GM-CSF	Biologend	MP1-22E9	505406	B196270	mouse	ICFC
IFNγ	Biologend	XMG1.2	505806	B232733	mouse	ICFC
CD45	Biologend	104	109823	N/A	mouse	ICFC
CD44	BD Biosciences	IM7	553133	53280	mouse	FC
CD25	Biologend	PC61.5	102008	B122202	mouse	FC
IL-4	eBioscience	11B11	17-7041-82	B138607	mouse	FC
Foxp3	eBioscience	FJK-16s	17-5773-82	E07303-1633	bovine, dog, cat, mouse, pig, rat	FC
Linage	Biologend	145-2c11,rb6-8c5,m1/70,ra3-6b2	133301	B142826	mouse	FC
CD127	Biologend	A7R34	135014	B142788	mouse	FC
CD117	Biologend	2B8	105812	B217855	mouse	FC
RORgt	eBioscience	B2D	12-6988-80	E02046-325	human, mouse, rhesus monkey	FC
TCRb	eBioscience	H57-597	553172	7047663	mouse	FC
RORgt	BD Bioscience	Q31-378	562663	3302666	mouse	FC, IHC, WB
beta-actin	Santa Cruz Biotechnology	AC-15	SC-8422	OAAD00224	human, mouse, rat	WB, IHC, IP
GFP	Life technology	polyclonal	A11122	1691382	tag	IHC, WB, ICC, IHC, IP, ChIP, FC, ELISA, IF
HA	Sigma-aldrich	114-2C-7	05-902R	53747	tag	WB
SRC1	Cell Signaling	128E7	2191S	1130X208	human, mouse, rat, monkey	WB, IP, IHC, ChIP
FLAG	Sigma-aldrich	M2	F1840	SLBQ6349V	tag	WB, IP, IHC, IF, ChIP

10. Eukaryotic cell lines

a. State the source of each eukaryotic cell line used.

HEK293t,Platinum-Eco

b. Describe the method of cell line authentication used.

Obtained from ATCC

c. Report whether the cell lines were tested for mycoplasma contamination.

All cells lines were negative for mycoplasma contamination.

d. If any of the cell lines used in the paper are listed in the database of commonly misidentified cell lines maintained by [ICLAC](#), provide a scientific rationale for their use.

No commonly misidentified cell lines were used.

► Animals and human research participants

Policy information about [studies involving animals](#); when reporting animal research, follow the [ARRIVE guidelines](#)

11. Description of research animals

Provide details on animals and/or animal-derived materials used in the study.

C57BL/6 mice, Rorc2 KO mice, Rorc2 mutant mice and Rag1 KO mice, female, 6-12weeks of age were used.

Policy information about [studies involving human research participants](#)

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

This study did not involve human research participants.